Effect of dietary calcium on cadmium absorption and retention in suckling rats

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Abstract

The effect of calcium supplementation on absorption and retention of cadmium in the suckling period was evaluated in Wistar rat pups of both sexes. Animals were maintained in the litters with the mother rats and supplemented with 1%, 3% or 6% calcium (as $CaHPO_4 \times 2H_2O$) in cow's milk by artificial feeding from day of birth 6 through 14. All rats were exposed to cadmium (as $CdCl_2 \times H_2O$) either orally or parenterally. Oral cadmium dose of 0.5 mg/kg body weight a day was administered through nine-day period of calcium supplementation and parenteral cadmium dose was injected subcutaneously in a single dose of 0.5 mg Cd/kg body weight prior to calcium supplementation. On experimental day 10 (at the age of pups of 15 days) all animals were killed and the liver, kidneys, brain and carcass (body without organs and skin) were removed for element analyses. Cadmium and essential elements calcium, zinc and iron were analysed in the tissues by atomic absorption spectrometry. Results showed that after oral exposure cadmium concentrations in all calcium-supplemented groups were significantly decreased in the organs and carcass and that the effect was dose-related. No such effect of calcium was found after parenteral cadmium exposure. Calcium supplementation perse significantly increased calcium concentration in the carcass and had no effect on iron in organs and zinc in carcass. It was concluded that calcium supplementation during the suckling period could be an efficient way of reducing oral cadmium absorption and retention without affecting tissue essential trace element concentrations.

Infants and young children are orally exposed to cadmium from food, water, household dust and/or garden soil (hand-to-mouth route of exposure) (IPCS 1992; Järup *et al.* 1998). Due to specific features of metal metabolism, the very young organism is at a higher risk for health effects of toxic metals such as cadmium than adults at the same level of environmental exposure (Kello & Kostial 1977; Kostial 1983; Kostial *et al.* 1991; Oskarsson *et al.* 1998).

Beside age, intestinal absorption of cadmium is influenced by a variety of factors including its chemical form, dose and route of exposure, environmental

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matrix in which it is contained, intestinal content, diet composition, nutritional status, and interactions of cadmium with other nutrients (Kello et al. 1979; IPCS 1992; WHO 1996; Diamond et al. 1998; Bhattacharyya et al. 2000). Recent work of Eklund & Oskarsson (1999) confirms that the composition of food has a great influence on the estimated mean weekly intake of dietary cadmium in infants. Soyabased formulas and diets with cereal products have a significantly higher content of cadmium than cow's milk formulas. It has been shown that essential minerals such as iron, calcium, phosphorus and zinc alone or in combination interact with intestinal (mainly duodenal) cadmium absorption and toxicity (Groten et al. 1991; IPCS 1992; Goyer 1995; Peraza et al. 1998). By increasing the calcium content in the diet, decreased cadmium absorption and retention in the body has been found. On the other hand, deficient dietary intake of calcium causes increased intestinal absorption of cadmium and its increased organ accumulation finally leading to enhancement of cadmium toxicity. Protective effects of calcium against cadmium absorption, accumulation and toxicity have been documented (reviewed by Brzóska & Moniuszko-Jakoniuk 1998).

Data on calcium and cadmium interactions in very young are not available. It is known that during the suckling period gastrointestinal absorption of mineral is very high due to increased nutritional requirements during the period of rapid growth and development (Kostial 1983; WHO 1996). This applies not only to the essential minerals such as calcium, but also to the toxic metals such as cadmium or lead. In our previous investigation we evaluated the influence of calcium supplementation on lead absorption, retention and elimination in suckling rats. It was found that supplementation with calcium hydrogen phosphate by artificial feeding during concomitant oral lead exposure significantly reduced body burden of lead (Varnai 1999; Varnai et al. 2000). In the present investigation we applied the same experimental model to assess cadmium absorption and retention in suckling rats exposed to cadmium either orally or parenterally under condition of oral calcium supplementation.

Materials and methods

Animals

Suckling Wistar rats of both sexes (from the Institute's breeding farm), six-day-old, with average weight of 14 g were used. Two days after birth five litters were reduced to eight pups each. During experiments pups were kept in litters with their mothers in individual policarbonate cages ($26 \times 20 \times 14$ cm; Ehret, Germany). Pups' body weights were recorded daily. The pine shaving beddings were changed every day.

The research procedures were performed according to the national law on the care and use of laboratory animals and were approved by the Croatian Ministry of Agriculture and Forestry.

Experimental design

Two experiments were performed. Experiment 1 was carried out to assess whether calcium supplementation influences intestinal cadmium absorption and/or incorporated cadmium elimination. Experiment 2 was

carried out to evaluate dose-relationship of the effect of calcium supplementation during oral cadmium exposure. In both experiments pups were exposed to cadmium and randomly designated to either control (unsupplemented) or calcium-supplemented group within each litter.

Calcium supplementation.

Calcium (as CaHPO₄×2H₂O, p.a. grade, Kemika Co., Zagreb, Croatia) was supplemented to cow's milk (commercial dairy product with 2.8% fat, 1200 mg/l calcium, 0.3 µg/l cadmium, 1 mg/l iron and 5 mg/l zinc) with addition of 6% calcium (Experiment 1) or increasing doses of 1%, 3% or 6% calcium (Experiment 2). Control (unsupplemented) group received cow's milk. Milk and calcium suspensions were administered to the pups during nine consecutive days starting on day of birth 6. Artificial feeding with a dropper was applied. The method closely resembles bottle-feeding of the infants and it has been originally introduced by Kostial et al. (1971). Drop-by-drop feeding was carried out with an automatic pipettor (25 μ l) during 7 h (9:00 a.m. till 4:00 p.m. with pauses). Number of drops of milk (in control, unsupplemented pups) or milk calcium-suspension (in supplemented pups) was increasing gradually by pups' age starting with 10 and ending with 45 drops per day. After 4:00 p.m. pups were returned to the mother rats where they remained the rest of the day and over night.

Cadmium exposure.

In both experiments pups were exposed to cadmium as cadmium chloride solution dissolved in distilled water when given orally and in saline when given subcutaneously ($CdCl_2 \times H_2O$, p.a. grade, Kemika Co., Zagreb, Croatia).

Experiment 1

There were four groups with 10 pups per group (2 pups per group in each of 5 litters):

Cd p.o. - orally Cd-exposed control

(unsupplemented) group;

Cd p.o. + Ca 6% - orally Cd-exposed and 6%

Ca-supplemented group;

Cd s.c. - subcutaneously Cd-exposed

control (unsupplemented) group;

Cd s.c + Ca 6% - subcutaneously Cd-exposed and

6% Ca-supplemented group.

In the first two groups cadmium was administered orally during the nine-day period of artificial feeding

(with cow's milk or with 6% calcium suspension in cow's milk) at a daily dose of 0.5 mg Cd/kg body weight. Total dose of cadmium was 4.5 mg Cd/kg body weight. Other two groups were administered a single subcutaneous injection of 0.5 mg Cd/kg body weight on 5th day of life, 24 h prior to beginning of calcium supplementation.

Experiment 2

There were four groups with 10 pups per group (2 pups per group in each of 5 litters):

Cd p.o. - orally Cd-exposed control (unsupplemented) group;

Cd p.o. + Ca 1% - orally Cd-exposed and 1% Ca-supplemented group;

Cd p.o. + Ca 3% - orally Cd-exposed and 3%

Ca-supplemented group;

Cd p.o. + Ca 6% - orally Cd-exposed and 6% Ca-supplemented group.

In all groups cadmium was administered orally in increasing doses (as 1%, 3% or 6% calcium suspension) during the nine-day period of artificial feeding in cow's milk at a daily dose of 0.5 mg/kg Cd kg body weight (total dose 4.5 mg Cd/kg body weight).

Sampling and element analysis

On the 10th day of the experiment (day of birth 15), rats were weighed and killed by exsanguination from the abdominal aorta in ether anaesthesia. In both experiments, the liver, both kidneys and brain, and in Experiment 1 also carcass (whole body without internal organs, entire gastrointestinal tract and skin) were removed for element analyses.

Fresh organ and carcass weights were measured. Samples were dried at 105 °C for 24 h, weighted and ashed at 450 °C in muffle furnace (Gallenkamp, UK) for 24 h (Blanuša & Breški 1981). After cooling down the samples were weighed again and adjusted to 10 ml with 10% HNO₃. All organ weights and carcass wet, dry and ash weights were recorded. Cadmium in the liver, iron in soft tissues and calcium and zinc in the carcass were analysed by flame atomic absorption spectrometry (AAS) on Varian instrument AA-375 (Varian; Australia). Cadmium in kidney and brain was analysed by electrothermal AAS on Varian AA300 with GTA 96. Deuterium correction was applied during all measurements.

To verify our chemical analysis Certified Standard Reference Material Bovine Liver 1577b (NIST, USA) and Animal Bone H-5 (IAEA, Austria) were analysed by the same procedure as the samples from the experiments. For cadmium, iron and zinc in Bovine Liver obtained values were 0.55, 184 and 127 μ g/g (certified values are 0.50, 184 and 127), respectively. For calcium and cadmium in Animal Bone obtained values were 217 mg/g and 0.026 μ g/g (reference values 212 and 0.023), respectively.

Statistical analysis

Results are presented as arithmetic means and standard errors of the means. Trace elements cadmium, iron and zinc were expressed as $\mu g/g$ tissue wet weight, whereas calcium in carcass was presented in mg g wet weight. In the first experiment statistical difference between the control and supplemented group was evaluated by Student's t-test (at the level of significance P < 0.05). The statistical differences between groups in the second experiment were analysed by general ANOVA/MANOVA with calcium supplementation as an independent variable. Where main effect of calcium supplementation was found, post hoc Duncan's multiple range test (at the level of significance P < 0.05) was applied. Statistica for Windows program (StatSoft release 5) was used for all statistical analyses.

Results

General effects

In both experiments body weight gain was about 2 g a day. At the end of both experiments average pup body weight was 33.0 g with no significant differences between the groups. No changes due to calcium supplementation were observed either in organ weights (wet or dry) or carcass weights (wet, dry or ash weight) (Table 1).

Effects on tissue cadmium concentration

We found that calcium supplementation during ongoing oral cadmium exposure caused a statistically significant reduction in tissue cadmium concentrations (Experiment 1). Cadmium concentration in the carcass was reduced to 57% of control values (Table 2), and the liver, kidneys and brain cadmium concentrations were reduced to 30%, 52% and 42% of control values (Table 3). By parenteral cadmium administration, calcium supplementation had no effect on cadmium

Table 1. Whole body weights, fresh and dry organ weights and wet, dry and ash carcass weights (g) in suckling rats^a.

Group	Body weight	Liver weight		Kidney weight		Brain weight		Carcass weight		
		Wet	Dry	Wet Dry		Wet	Dry	Wet	Dry	Ash
Experiment 1										
Cd p.o.	34.5 ± 1.39	$1.13 \pm 0.046 \ 0.271 \pm 0.011$	0.271 ± 0.011	$0.408 \pm 0.021 \ 0.075 \pm 0.004$	5 ± 0.004	1.33 ± 0.030	$1.33 \pm 0.030 \ \ 0.198 \pm 0.005$	$14.7 \pm 0.649 \ \ 3.71 \pm 0.158 \ \ 0.643 \pm 0.032$	3.71 ± 0.158	0.643 ± 0.032
Cd p.o. + Ca 33.4 ± 1.17 6%	33.4 ± 1.17	$1.07 \pm 0.041 \ 0.259 \pm 0.010$	0.259 ± 0.010	$0.375 \pm 0.015 \ 0.070 \pm 0.003$	0 ± 0.003	1.26 ± 0.037	$1.26 \pm 0.037 0.188 \pm 0.006$	$13.9 \pm 0.362 \ \ 3.57 \pm 0.095 \ \ 0.669 \pm 0.025$	5.57 ± 0.095	0.669 ± 0.025
Cd s.c.	34.1 ± 1.16	1.09 ± 0.052	0.256 ± 0.012	$0.384 \pm 0.018 \ 0.07$	0.070 ± 0.003	1.29 ± 0.035	$1.29 \pm 0.035 \ 0.194 \pm 0.006$	$14.3 \pm 0.600 \ \ 3.61 \pm 0.156 \ \ 0.640 \pm 0.030$	6.61 ± 0.156	0.640 ± 0.030
Cd s.c. + Ca 6% 32.7 ± 1.14	32.7 ± 1.14	$1.10 \pm 0.041 \ 0.258 \pm 0.010$	0.258 ± 0.010	$0.389 \pm 0.015 \ 0.074 \pm 0.003$	4 ± 0.003	1.29 ± 0.018	$1.29 \pm 0.018 \ 0.198 \pm 0.003$	13.8 ± 0.520 3.58 ± 0.142 0.679 ± 0.025	0.58 ± 0.142	0.679 ± 0.025
Experiment 2										
Cd p.o.	33.7 ± 0.559	$1.09 \pm 0.031 0.269 \pm 0.022$	0.269 ± 0.022	$0.337 \pm 0.008 \ 0.065 \pm 0.002$	5 ± 0.002	1.22 ± 0.032	$1.22 \pm 0.032 0.190 \pm 0.005$	$14.7 \pm 0.289 \ \ 3.81 \pm 0.122 \ \ 0.600 \pm 0.009$	3.81 ± 0.122	0.600 ± 0.009
Cd p.o. + Ca 1% 32.9 ± 0.607	32.9 ± 0.607	1.07 ± 0.032	0.257 ± 0.027	$0.340 \pm 0.008 \ 0.063 \pm 0.002$	3 ± 0.002	1.20 ± 0.021	$1.20 \pm 0.021 0.188 \pm 0.003$	$14.4 \pm 0.393 \ \ 3.78 \pm 0.152 \ \ 0.610 \pm 0.003$	3.78 ± 0.152	0.610 ± 0.003
Cd p.o. + Ca 3% 32.5 ± 0.515	32.5 ± 0.515	$1.05 \pm 0.019 \ \ 0.259 \pm 0.020$	0.259 ± 0.020	0.330 ± 0.007 0.06	0.062 ± 0.002	1.24 ± 0.024	$1.24 \pm 0.024 \ \ 0.193 \pm 0.004$	$13.9 \pm 0.275 \ \ 3.60 \pm 0.096 \ \ 0.621 \pm 0.010$	960.0 ± 099	0.621 ± 0.010
Cd p.o. + Ca 6% 32.7 ± 0.565	32.7 ± 0.565	$1.07 \pm 0.026 \ 0.266 \pm 0.017$	0.266 ± 0.017	$0.333 \pm 0.008 \ 0.062 \pm 0.001$	2 ± 0.001	1.21 ± 0.025	$1.21 \pm 0.025 \ 0.188 \pm 0.004$	14.1 ± 0.271 3.67 ± 0.083 0.637 ± 0.009	67 ± 0.083	0.637 ± 0.009

^aSix-day-old suckling rats received cadmium (as $CdCl_2 \times H_2O$ in distilled water) either per os (Cd p.o.) in totalamount of 4.5 mg Cd/kg body weight during 9 days with daily amount of 0.5 mg Cd/kg body weight divided into two drops per day (Experiment 1 and Experiment 2) or subcutaneously (as $CdCl_2 \times H_2O$ in saline) as a single dose of 0.5 mg Cd/kg body weight (Cd.s.c.)prior to the 9-day experiment (Experiment 1). Calcium supplement (as CaHPO₄x2H₂O) was given in 6% Ca suspension (Experiment 1) or in increasing concentrations of 1%, 3% or 6% Ca (Experiment 2) in cow's milk by artificial

feeding during 7 h a day over 9 days. Results are presented as arithmetic means \pm SEM of 10 animals per group.

Table 2. Concentrations of cadmium, zinc and calcium in carcass of suckling rats orally supplemented with 6% calcium in cow's milk and orally or subcutaneously exposed to cadmium#.

Group	Element concentrations in carcass						
	Cadmium μg/g wet wt)		Zinc	Calcium (mg/g wet wt)			
Experiment 1							
Cd p.o.	0.044 ± 0.006		28.1 ± 0.202	14.6 ± 0.195			
Cd p.o. + Ca 6%	$0.024 \pm 0.001^*$	(57)	29.1 ± 0.554	$16.1 \pm 0.440^*$	(110)		
Cd s.c.	0.102 ± 0.010		29.8 ± 0.464	13.8 ± 0.674			
Cd s.c. + Ca 6%	0.090 ± 0.008	(88)	30.0 ± 0.669	$16.9 \pm 0.347^*$	(123)		

^{*}Experimental protocol same as described under Table 1.

Results are presented as arithmetic means \pm SEM of 10 animals per group (percentage of value in respective unsupplemented control group in parentheses). *Significantly different from respective control group at P < 0.05 by Student's t-test

Table 3. Concentrations of cadmium and iron in the liver, kidneys and brain of suckling rats ($\mu g/g$ wet organ weight) orally supplemented with 1%, 3% or 6% calcium in cow's milk and orally or subcutaneously exposed to cadmium#.

Group						
	Liver		Kidneys		Brain	
	C	admium co	ncentrations (μg/g wet	wt)		
Experiment 1						
Cd p.o.	1.37 ± 0.192		1.04 ± 0.110		0.024 ± 0.004	
Cd p.o. + Ca 6%	$0.406 \pm 0.033^*$	(30)	$0.544 \pm 0.037^*$	(52)	$0.010 \pm 0.002^*$	(42)
Cd s.c.	3.42 ± 0.323		0.745 ± 0.060		0.033 ± 0.005	
Cd s.c. + Ca 6%	2.95 ± 0.231	(86)	0.588 ± 0.059	(79)	0.037 ± 0.006	(119)
Experiment 2						
Cd p.o.	1.17 ± 0.067^{a}		1.09 ± 0.063^{a}		0.014 ± 0.002^{ab}	
Cd p.o. + Ca 1%	0.974 ± 0.069^{b}	(83)	0.910 ± 0.057^{b}	(84)	0.019 ± 0.004^{a}	(136)
Cd p.o. + Ca 3%	0.582 ± 0.026^{c}	(50)	0.679 ± 0.033^{c}	(63)	0.011 ± 0.003^{bc}	(79)
Cd p.o. + Ca 6%	$0.403 \pm 0.034^{\mathrm{d}}$	(34)	0.565 ± 0.056^{c}	(52)	0.005 ± 0.001^{c}	(36)
		Iron conce	entrations (μg/g wet wi	t)		
Experiment 1						
Cd p.o.	42.8 ± 2.22		27.9 ± 1.62		11.1 ± 0.766	
Cd p.o. + Ca 6%	43.1 ± 2.76		24.7 ± 1.56		9.76 ± 0.733	
Cd s.c.	47.9 ± 5.00		26.9 ± 0.80		10.4 ± 0.485	
Cd s.c. + Ca 6%	46.2 ± 3.66		26.6 ± 1.20		9.69 ± 0.806	
Experiment 2						
Cd p.o.	46.9 ± 4.45		26.7 ± 1.14		10.6 ± 0.59	
Cd p.o. + Ca 1%	40.6 ± 1.45		24.9 ± 0.80		10.5 ± 0.44	
Cd p.o. + Ca 3%	40.4 ± 3.68		25.6 ± 0.80		10.2 ± 0.37	
Cd p.o. + Ca 6%	41.5 ± 2.70		24.4 ± 0.88		9.94 ± 0.25	

^{*}Experimental protocol same as described under Table 1.

Results are presented as arithmetic means \pm SEM of 10 animals per group (percentage of value in respective unsupplemented control group in parentheses).

^{*}Significantly different from respective control value at P<0.05 by Student's t-test (Experiment 1). abcd Significant differences between groups indicated by different superscript letters at P < 0.05 by Duncan's multiple range test (Experiment 2).

tissue concentrations either in carcass (Table 2) or in organs (Table 3).

In Experiment 2, by oral cadmium exposure and the highest dose of calcium (6% Ca) supplementation, cadmium concentrations in organs were the same as in Experiment 1 at the same dose of calcium supplementation (carcass was not analysed). At the 3% calcium supplementation, cadmium concentrations in the liver, kidneys and brain were significantly reduced to 50%, 63% and 79% of control values (Table 3). At the lowest level of calcium (1% Ca) supplementation, significant reduction in cadmium concentrations were observed in the liver and kidneys (to about 80% of control values) and there was no effect in the brain.

Effects on tissue essential elements

By calcium supplementation no change in carcass zinc concentration was found. Carcass calcium concentrations were significantly increased in all calcium-supplemented pups (Experiment 1, Table 2).

In both experiments calcium supplementation caused no changes in iron concentrations in the liver, kidneys or brain irrespective of the route of cadmium exposure (Table 3).

Discussion

Several authors (e.g., Kozlowska et al. 1993; Rimbach et al. 1995) have found adverse effect of oral cadmium exposure on growth and survival in rats. Oral and subcutaneous cadmium doses chosen in our experiments were much lower than respective LD₅₀ doses reported for suckling animals (Kostial et al. 1979). Daily oral dose was 3% of LD₅₀ determined for oral exposure and subcutaneous dose was 10% of LD₅₀ determined for intraperitoneal exposure to cadmium in suckling rats. At these cadmium doses pups gained about 2 g body weight per day and their general appearance (mobility, reaction to mechanical stimuli) and expected developmental changes (fur growth, eye and ear opening, development of motoric abilities and increase of the ability to swallow drops) were not affected.

The quantity of calcium supplementation in our experiments was chosen after literature search of data on calcium concentration in cow's and rat's milk. Most of the reported results for calcium concentration in mature cow's milk are between 1.1 and 1.3 g/kg (IAEA 1982). Taking an average of the reported data for rat's

milk intake and its calcium concentration, we have estimated the quantity of ingested calcium from mother rat's milk during experiment to be about 100 mg plus 8 mg calcium through cow's milk (Luckey *et al.* 1954; Kametaka *et al.* 1974; Yagil *et al.* 1976; Keen *et al.* 1981; Auestad *et al.* 1989; Nicholas & Hartmann 1991). Calcium supplementations of 1%, 3% and 6% calcium suspension in cow's milk during 9 days contained 76, 211 and 415 mg calcium. That is, by giving 6% calcium supplementation we increased calcium intake about 4 times.

It has been previously found that calcium-deficient diet enhances cadmium absorption and that cadmium has a direct effect on bone in mice and rats (Wang & Bhattacharyya 1993; Whelton et al. 1997a, b; Piasek et al. 1997). In our previous investigations we found that adult rats on lower calcium diet and concomitantly exposed to cadmium in drinking water had reduced mineral content in the bone, increased cadmium concentration in the liver, and changed zinc and iron tissue concentrations in comparison with the rats on higher calcium diet (Blanuša et al. 1983; Piasek et al. 1997). Studies with calcium supplementation in adult rats showed lower whole body retention of cadmium with increasing dietary calcium level (Kello et al. 1979). In growing rats the best reduction of cadmium accumulation and toxicity has been obtained with mineral composition supplement containing iron, calcium/phosphorus and zinc (Groten et al. 1991; Walter et al. 2000). Results of our present investigation on suckling rats show that calcium supplementation primarily influences absorption and retention of orally administered cadmium and has no effect on elimination of incorporated cadmium. In addition, our results show that the effect of calcium supplementation on decreasing tissue concentration of toxic metal in young organism is dose-related. That is, after oral cadmium exposure, even the lowest level of calcium supplementation caused a significant reduction of cadmium retention in pups' liver and kidneys.

From presented results it might be concluded that calcium supplemented milk could be a way to reduce cadmium tissue levels early in life. However, before any final conclusions on the effectiveness of such preventive treatment, more data are needed to evaluate the health benefits or risks of calcium supplementation at this early period of life.

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References

- Auestad N, Korsak RA, Bergstrom JD, Edmond J. 1989 Milk-substitutes comparable to rat's milk; their preparation, composition and impact on development and metabolism in the artificially reared rat. *Br J Nutr* **61**, 495–518.
- Bhattacharyya MH, Wilson AK, Rajan SS, Jonah M. 2000 Biochemical pathways in cadmium toxicity. In: Zalups RK, Koropatnick J, eds. *Molecular Biology and Toxicology of Metals*. London-New York: Taylor & Francis; 34–74.
- Blanuša M, Breški Dj. 1981 Comparison of dry and wet ashing procedures for cadmium and iron determination in biological material by atomic-absorption spectrophotometry. *Talanta* 28, 681–684
- Blanuša M, Rabar I, Kostial K. 1983 The health effect of cadmium in relation to changes in physiological levels of dietary calcium and iron in rats. In: Brätter P, Schramel P, eds. *Trace Element-Analytical Chemistry in Medicine and Biology*. Vol 2. Berlin-New York: Walter de Gruyter & Co; 401–411.
- Brzóska MM, Moniuszko-Jakoniuk J. 1998 The influence of calcium content in diet on cumulation and toxicity of cadmium in the organism. *Arch Toxicol* **72**, 63–73.
- Diamond GL, Goodrum PE, Felter SP, Ruoff WL. 1998 Gastrointestinal absorption of metals: a review. *Drug Chem Toxicol* 21, 223–251
- Eklund G, Oskarsson A. 1999 Exposure of cadmium from infant formulas and weaning foods. Food Addit Contam 16, 509–519.
- Goyer RA. 1995 Nutrition and metal toxicity. *Am J Clin Nutr* **61**(suppl), 646S–650S.
- Groten JP, Sinkeldam EJ, Muyis T, Luten JB, van Bladeren PJ. 1991 Interaction of dietary Ca, P, Mg, Mn, Cu, Fe, Zn and Se with the accumulation and oral toxicity of cadmium in rats. *Fd Chem Toxic* 29, 248–258.
- International Atomic Energy Agency (IAEA). 1982 Elemental Composition of Human and Animal Milk. TECDOC-269. Vienna: IAEA.
- International Programme on Chemical Safety (IPCS). 1992 Cadmium. Environmental Health Criteria (EHC) 134. Geneva: WHO.
- Järup L, Berglund M, Elinder CG, Nordberg G, Vahter M. 1998 Health effects of cadmium exposure – a review of the literature and a risk estimate. *Scand J Work Environ Health* 24(Suppl 1), 1–52.
- Kametaka M, Inaba J, Ichikawa R. 1974 Estimation of the daily milk intake of the suckling rat using the turnover rate of potassium. J Nutr Sci Vitaminol 20, 421–429.
- Keen CL, Lönnerdal B, Clegg M, Hurley LS. 1981 Developmental changes in composition of rat milk: trace elements, minerals, protein, carbohydrate and fat. J Nutr 111, 226–235.
- Kello D, Kostial K. 1977 Influence of age and milk diet on cadmium absorption from the gut. Toxicol Appl Pharmacol 40, 277–282.
- Kello D, Dekanić D, Kostial K. 1979 Influence of sex and dietary calcium on intestinal cadmium absorption in rats. Arch Environ Health 34, 30–33.

- Kostial K, Šimonović I, Pišonić M. 1971 Lead absorption from the intestine in newborn rats. *Nature* 233, 564.
- Kostial K, Kello D, Blanuša M, Maljković T, Rabar I. 1979 Influence of some factors on cadmium pharmacokinetics and toxicity. Environ Health Perspect 28, 89–95.
- Kostial K. 1983 The absorption of heavy metals by the growing organism: experimental experience with animals. In: Schmidt EHF, Hildebrandt AG, eds. *Health Evaluation of Heavy Metals*. Berlin-Heidelberg: Springer-Verlag; 99–104.
- Kostial K, Blanuša M, Maljković T, Kargačin B, Piasek M, Momčilović B, Kello D. 1991 Age and sex infuence the metabolism and toxicity of metals. In: Momčilović B, ed. *Trace Elements in Man and Animals TEMA-7*. Zagreb, Croatia: IMI; 11/1–11/5.
- Kozlowska K, Brzozowska A, Sulkowska J, Roszkowski W. 1993 The effect of cadmium on iron metabolism in rats. *Nutr Res* 13, 1163–1172.
- Luckey TD, Mende TJ, Pleasants J. 1954 The physical and chemical characterization of rat's milk. *J Nutr* **54**, 345–359.
- Matek M, Blanuša M, Varnai VM, Piasek M, Kostial K. 2000 Effect of dietary calcium on cadmium metabolism in suckling rats. In: 2nd Croatian Congress of Toxicology with International Participation, Pula, Croatia. Final Programme and Abstract Book, 52.
- Nicholas KR, Hartmann PE. 1991 Milk secretion in the rat: Progressive changes in milk composition during lactation and weaning and the effect of diet. Comp Biochem Physiol 98A, 535–542.
- Oskarsson A, Palminger Hallén I, Sundberg J, Petersson Grawé KP. 1998 Risk assessment in relation to neonatal metal exposure. Analyst 123, 19–23.
- Peraza MA, Ayala-Fierro F, Barber DS, Casarez E, Rael LT. 1998 Effects of micronutrients on metal toxicity. *Environ Health Perspect* 106, 203–216.
- Piasek M, Schönwald N, Blanuša M, Kostial K, Momčilović B. 1997 Effect of cadmium exposure and diet with lower calcium on bone in female rats. In: Fischer PWF, L'Abbé MR Cockell KA, Gibson RS, eds. *Trace Elements in Man and Animals – 9*. Ottawa, Canada: NRC Research Press; 527–528.
- Rimbach G, Brandt K, Most E, Pallauf J. 1995 Supplemental phytic acid and microbial phytase change zinc bioavailability and cadmium accumulation in growing rats. *J Trace Elements Med Biol* 9, 117–122.
- Varnai VM. 1999 The effect of calcium on lead retention in rats. M.Sc. Thesis. Zagreb: University of Zagreb.
- Varnai VM, Piasek M, Blanuša M, Matek M, Kostial K. 2000 Calcium supplementation reduces lead absorption in the gut during suckling period In: 2nd Croatian Congress of Toxicology with International Participation, Pula, Croatia. Final Programme and Abstract Book. 51.
- Walter A, Rimbach G, Most E, Pallauf J. 2000 Effect of calcium supplements to a maize-soya diet on the bioavailability of minerals and trace elements and the accumulation of heavy metals in growing rats. J Vet Med A 47, 367–377.
- Wang C, Bhattacharyya MH. 1993 Effect of cadmium on bone calcium and ⁴⁵Ca in nonpregnant mice on a calcium-deficient diet: Evidence of direct effect of cadmium on bone. *Toxicol Appl Pharmacol* 120, 228–239.
- Whelton BD, Peterson DP, Moretti ES, Dare H, Bhattacharyya MH. 1997a Skeletal changes in multiparous, nulliparous and ovariectomized mice fed either a nutrient-sufficient or deficient diet containing cadmium. *Toxicology* 119, 103–121.
- Whelton BD, Peterson DP, Moretti ES, Mauser RW, Bhattacharyya MH. 1997b Kidney changes in multiparous, nulliparous and

ovariectomized mice fed either a nutrient-sufficient or -deficient diet containing cadmium. *Toxicology* **119**, 123–140.

World Health Organization (WHO). 1996 Trace Elements in Human Nutrition and Health. Geneva: WHO.

Yagil R, Etzion Z, Berlyne GM. 1976 Changes in rat milk quantity and quality due to variations in litter size and high ambient temperature. *Lab Anim Sci* **26**, 33–37.